



Research Article

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COMPARATIVE INHIBITORY POTENTIAL OF ASHODHITA (RAW) AND SHODHITA *CURCUMA LONGA* LINN ON α -AMYLASE AND α -GLUCOSIDASE

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ABSTRACT

Turmeric (Haridra) is one of the versatile herb for spices, condiment and anti-diabetic activity in Indian System of medicine and tropical country. To compare the in-vitro enzymatic inhibitory effect (α -amylase and α -glucosidase) for supporting purified (shodhita) and raw turmeric samples. Authentic unprocessed and processed (ayurvedic classic) turmeric powder extracts prepared by maceration with methanol and subjected (in different level concentration) to the anti-hyperglycemic selective enzymatic activity and colour reaction was measured by UV-VIS spectroscopy. This study proves the alteration in the inhibitory effect of raw and shodhita samples of Haridra on α -amylase of IC_{50} in decreasing order TZ (113.72) < TW (119.22) < TR (121.861) < TT (161.35) < TGM (218.85) and α -glucosidase TZ (121.32) < TW (139.48) < TR (145.29) < TT (185.51) < TGM (276.12) respectively. Purified turmeric samples gives better effect than raw sample. In that way shodhan effect established indirectly synergistic and supra-additive effect of turmeric in context to antidiabetic.

Keywords: Turmeric, Shodhana, α -amylase, α -glucosidase, antidiabetic

INTRODUCTION

Indian medicinal plants used in the Ayurvedic traditional system to treat diabetes are more valuable source of novel anti-diabetic agents now a days due to less expensive when compared to synthetic anti-hyperglycemic agents and have very less side effect.^{1,2} It will be assessed that more than 200 species about plants show hyperglycaemic properties.^{3,4,5,6} Turmeric (*Curcuma longa* Linn) belongs to the family Zingiberaceae, is also referred to Ayurvedic system of medicine and also found good anti-diabetic effect as it is describe in various classical texts^{7,8,9} as well as modern texts^{10,11} and research paper^{12,13,14}. Shodhana is not just a purification process, it is a process of enhancement of drug quality as per Ayurveda.¹⁵ On the other hand the classical (Ayurvedic) principle drug act as a whole there is no need for isolation of primary and secondary plant metabolite due to belief in the holistic sense¹⁶ which counteract modern science selecting one –two marker to ignore complexity of herbal medicine¹⁷. Accepting the extraction technique of selected samples to find comparative effect of natural α -amylase and α -glucosidase inhibitors in quest of finding the comparative anti-diabetic efficacy of raw and all shodhita. These α -amylase and α -glucosidase inhibitory activity was conducted due to therapeutic approach to prevent postprandial hyperglycemia is to retard the digestion and absorption of carbohydrates in the gastrointestinal

tract through inhibition of enzymes such as α -amylase and α -glucosidase. The α -amylase (α -1, 4-glucan-4-glucanohydrolases) is one of the major secretory products of the pancreas and salivary glands, playing a role in digestion of starch and glycogen and can be found in microorganisms, plants and higher organisms. α -glucosidase enzyme acted on maltose, maltotriose and branched oligosaccharides of α -(1-6) and α -(1-4) oligoglucans to monosaccharaides and also degraded glucose absorption in the blood stream. On the other hand, α -amylase hydrolysed of starch of various type of low molecular weight saccharides¹⁸. Although, these activities for Turmeric have been done previously^{19, 20, 21} but none of them have done shodhan of Haridra which make this work noval and significant. In this study, methanolic extracts of ashodhita and shodhita Haridra powders have been tested for α -amylase and α -glucosidase inhibitory activities.

MATERIALS AND METHODS

Samples description

Standard Sample (AC)

Acarbose tablets IP 50 mg of brand name “Glucobay 50” strip was purchased from Jamnagar which having batch no. P16141 and manufactured by Bayer Pharmaceutical Pvt. Ltd. At village Malpur, Baddi- 1732015.

Test Samples

Sample (TR): Methanolic extract of powder of raw turmeric rhizome without treated any media.

Sample (TT): Methanolic extract of powder of turmeric rhizome after treated with Takra (Nimajjana of Turmeric mother rhizome in takra for 10 days²², it is prepared with 1/4th amount of water²³)

Sample (TW): Methanolic extract of powder of turmeric rhizome after treated water (Swedana/boiling of turmeric mother rhizome in drinking mineral water for 3 hours²⁴)

Sample (TGM): Methanolic extract of powder of turmeric rhizome after boiling in water for 3 hours followed by steaming with fresh Gomutra (Cow's urine)

Sample (TZ): Methanolic extract of powder of turmeric rhizome after boiling in fresh Gomutra (1hour) followed by boiling in Panchapallava Kwatha (1hour) followed by boiling in Mundi Kwatha (1hour), followed by steaming in fresh Gomutra (15 minutes)^{25, 26}

Activities

α -Amylase inhibitory activity

The α -amylase inhibitory activity was determined according to the method described by Miller. A total of 500 μ l of test samples and standard drug (50-200 μ g/ml) were added to 500 μ l of 0.20 mM phosphate buffer (pH 6.9) containing α -amylase 500 μ l (0.5mg/ml) solution and were incubated at 25°C for 10 min. After these, 500 μ l of a 1% starch solution followed by 0.02 M sodium phosphate buffer 500 μ l (pH 6.9) was added to each tube. The

reaction mixtures were then incubated at 25°C for 10 min. The reaction was stopped with 1.0 ml of 3, 5 dinitrosalicylic acid colour reagent. The test tubes were then, incubated in a boiling water bath for 5 min and cooled at room temperature. The reaction mixture was then diluted after adding 10 ml distilled water and absorbance was measured at 540 nm. Control represent 100% enzyme activity and were conducted in similar way by replacing extract with vehicle.^{27, 28}

α -Glucosidase inhibitory activity

The inhibitory activity was determined by incubating a solution of starch substrate (2 % w/v maltose or sucrose) 500 μ l with 1000 μ l (0.2 M) Tris buffer pH 8.0 and various concentration of plant extract (50 μ g to 200 μ g), keep it for 5 min at 37°C. The reaction was initiated by adding 1 ml of α -glucosidase enzyme (1U/ml) to it followed by incubation for 10 min at 37°C. Then, the reaction mixture was heated for 2 min in boiling water bath to stop the reaction and then standard glucose reagent was added 250 μ l in each test tube. The amount of liberated glucose is measured by glucose oxidase peroxidase method and absorbance was measured at 510 nm.^{29, 30, 31}

The results of both were expressed as % inhibition calculated using the formula:

$$\text{Inhibitory activity of } \alpha\text{-amylase enzyme} = \frac{(\text{Abs. of Test} - \text{Abs. of control}) \times 100}{\text{Abs. of Test}}$$

Table 1: α -Amylase and α -glucosidase inhibitory activity of methanolic extracts of Turmeric Test samples and Reference drug Acarbose

Samples	Concentration (in μ g/ml)	Percentage of Inhibitions	
		α -Amylase	α -Glucosidase
AC (Standard)	50	47.53	49.81
	100	53.01	53.47
	150	60.2	57.59
	200	62.5	62.58
TR	50	40	42.73
	100	49.13	45.01
	150	54.29	50.55
	200	58.06	54.88
TT	50	36.4	31.02
	100	42.23	40.56
	150	49.65	45.37
	200	54.11	51.28
TW	50	40.57	42.72
	100	49.65	46.91
	150	54.11	50.78
	200	58.33	54.89
TGM	50	26.41	34.17
	100	33.89	37.34
	150	40.45	40.22
	200	47.29	45.11
TZ	50	41.35	43.78
	100	50.79	49.63
	150	54.54	52.51
	200	58.51	55.17

Table 2: IC₅₀ of Reference drug Acarbose and Turmeric Test samples

Samples	IC ₅₀ (in µg)	
	In α - Amylase	In α - Glucosidase
AC (standard)	69.24 $y = 0.1042x + 42.785, R^2 = 0.9665$	48.66 $y = 0.0823x + 45.995, R^2 = 0.9905$
TR	121.86 $y = 0.1187x + 35.535, R^2 = 0.9591$	145.297 $y = 0.084x + 37.795, R^2 = 0.9773$
TT	161.35 $y = 0.1211x + 30.46, R^2 = 0.9919$	185.51 $y = 0.1312x + 25.66, R^2 = 0.9773$
TW	119.22 $y = 0.1155x + 36.23, R^2 = 0.9605$	139.48 $y = 0.0808x + 38.73, R^2 = 0.9998$
TGM	218.85 $y = 0.1384x + 19.71, R^2 = 0.9993$	276.12 $y = 0.0714x + 30.285, R^2 = 0.9845$
TZ	113.72 $y = 0.110x + 37.49, R^2 = 0.943$	121.32 $y = 0.0741x + 41.01, R^2 = 0.9592$

Note: IC₅₀ = Half maximal inhibitory concentration

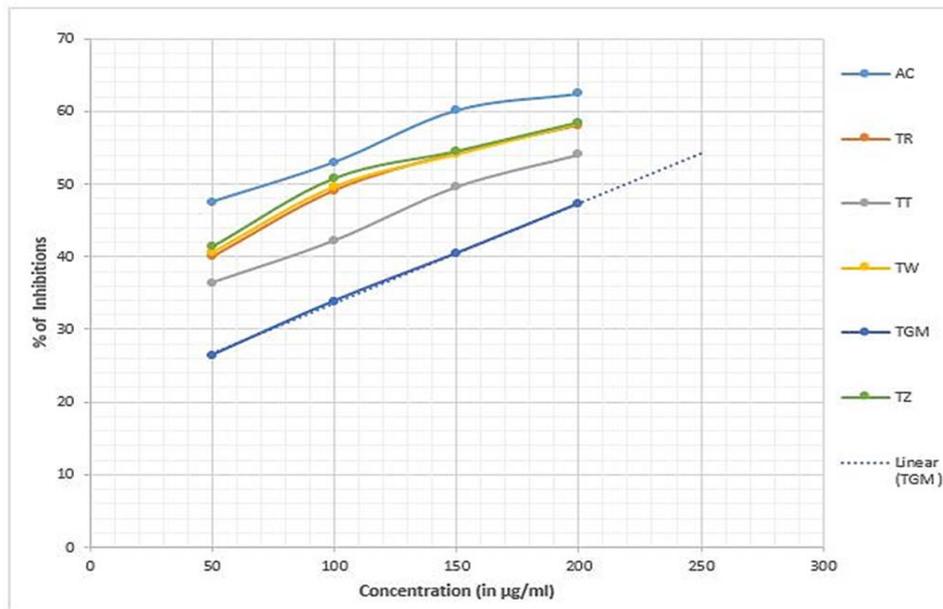


Figure 1: Graphical representation of Methanolic extracts of test samples, potential of inhibition of α-amylase enzyme.

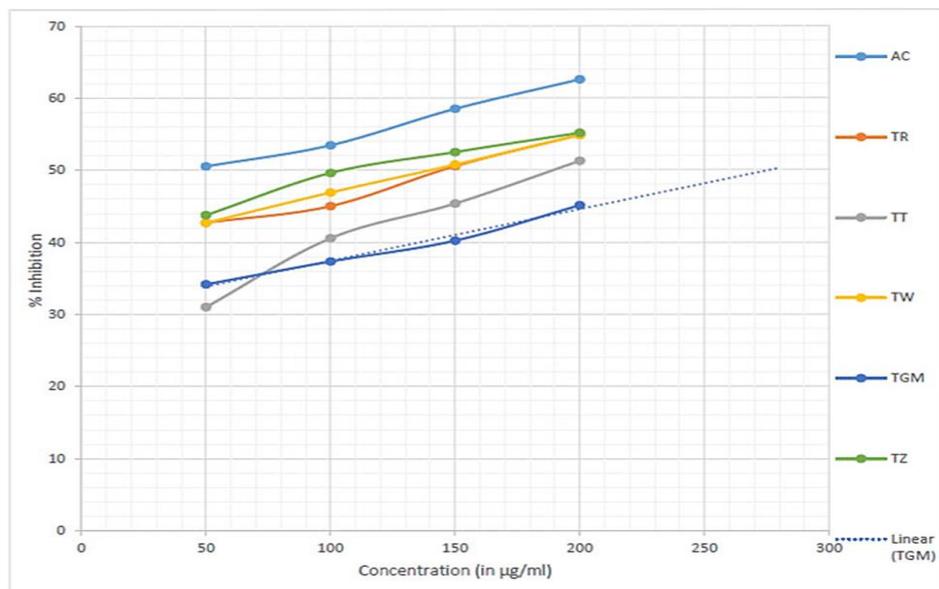


Figure 2: Graphical representation of Methanolic extracts of test samples' potential of inhibition of α-glucosidase enzyme.

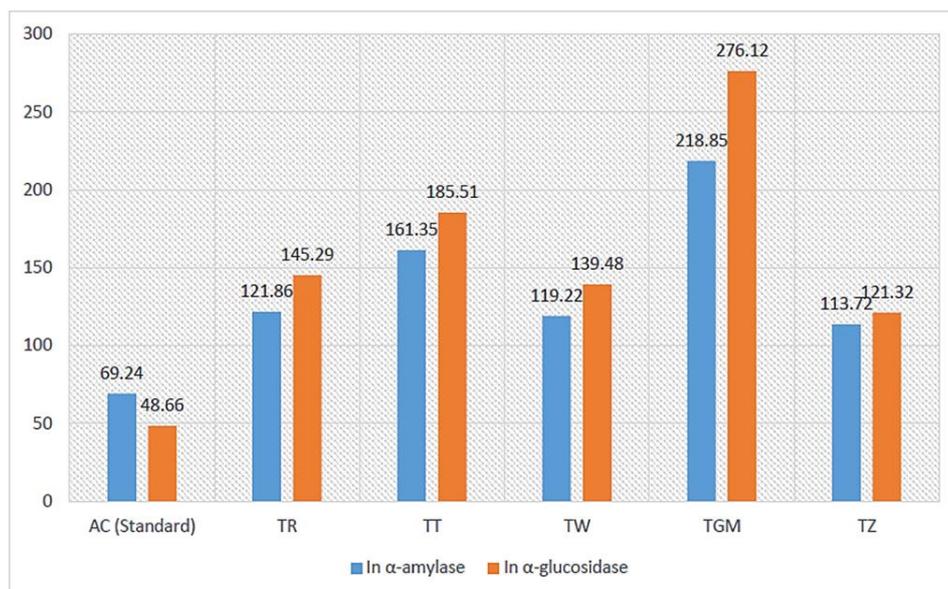


Figure 3: Comparison of IC_{50} dose of test samples for inhibition of α -amylase & α -glucosidase enzymatic activity

OBSERVATIONS AND RESULTS

There was a dose-dependent inhibitory effect of raw and Shodhita Haridra activity on α -amylase and α -glucosidase enzyme. It is evident from Table 3.1 that comparatively more inhibitory activity on α -amylase enzyme was demonstrated by standard drug Acarbose.

Test samples TR, TW and TZ have comparatively more inhibitory effect than other test samples like TT and TGM. Graphical representation shows similar pattern of inhibitory effect of TR, TW and TZ.

TGM showed lowest inhibition activity than other test samples, even it could not get IC_{50} . So, trendline in the graph was extended to 50 points, to get the IC_{50} . Same thing can be seen in case of α -glucosidase enzyme activity. Here, trendline of TGM was extended to 80 points to get the IC_{50} . From the above graphical representation of both α -amylase and α -glucosidase inhibition, TZ showed comparatively more inhibitory effect than others. Quantitatively, it's not a big value but comparatively, it is a significant difference.

IC_{50} of all samples and standard were calculated by using regression equation by linear curve drawn as trendline for both α -amylase and α -glucosidase. Although, IC_{50} of Acarbose in α -glucosidase could not be calculated because trendline has range 53.47% to 60.58 % and regression equation fail to give IC_{50} . Haridra samples show good effect in context to inhibition of α -amylase and α -glucosidase enzymes, however TZ show better inhibition than all other samples because it requires minimum dose to get IC_{50} . (Table 2)

DISCUSSION

In treatment of type II diabetes mellitus treatment concern key role control the human pancreatic α -amylase which increases in post-prandial glucose levels. Inhibitory activity of such type of enzyme, α -amylase in the form of dealing carbohydrate digestion, reduction of absorption blood glucose level^{32,33,34}. Recent research trend is focused on inhibition of carbohydrate metabolizing enzyme. Recently acarbose and miglitol such type of inhibitor are successfully used in clinically. To search naturally

occurring herbal samples which treated traditional shodhan process in extract level inhibitory activity of α -amylase and α -glucosidase. This study proves the alteration in the inhibitory effect of raw and Shodhita samples of Haridra on α -amylase and α -glucosidase. TZ show best IC_{50} in among all the samples then, followed by raw Haridra; TR and then, TW > TT > TGM in both enzymatic inhibition activity. (see in below cluster chart)

CONCLUSION

Pharmacological in-vitro enzymatic study was done to check the anti-diabetic property of raw and Shodhita Turmeric, in context to inhibitory activity of the α -amylase and α -glucosidase. Both methods were very sensitive with respect to time interval. Variation in the properties of inhibition of both enzymes can be seen as their decreasing order of IC_{50} dose (μg) in α -amylase: TZ (113.72) < TW (119.22) < TR (121.861) < TT (161.35) < TGM (218.85) and in α -glucosidase: TZ (121.32) < TW (139.48) < TR (145.29) < TT (185.51) < TGM (276.12). So, TZ is effective at minimum dose levels than others, shows significant therapeutic effect of Shodhana. Although TW and TR is also close to TZ but TZ is consider more better due to it shows minimum curcumin level in analytical section, that proves some special properties of media has increase the therapeutic property of TZ, which proves the synergic approach of Shodhana.

REFERENCES

1. Grover JK, Yadav S, Vats V: Medicinal plants of India with anti-diabetic potential. J Ethnopharmacol 2002, 81:81-100.
2. Mukherjee PK, Maiti K, Mukherjee K, Houghton PJ: Leads from Indian medicinal plants with hypoglycemic potentials. J Ethnopharmacol 2006, 106(1):1-28.
3. Jia W, Gao W, Tang L. Antidiabetic herbal drugs officially approved in China. Phytotherapy Research. 2003, 17(10):1127-34.
4. Satyavati G V, Raina M K & Sharma M, Medicinal plants of India, Vol 1, (Indian Council of Medical Research, New Delhi) 1976.
5. Nadkarni KM. Indian Medicinal Plants and Drugs with Their Medicinal Properties and Uses, 2006 edition, published by Srishti book distributors. (1910): 153-154.

6. Shukia R, Sharma SB, Puri D, Prabhu KM, Murthy PS. Medicinal plants for treatment of diabetes mellitus. Indian Journal of Clinical Biochemistry. 2000 Aug 1;15(1):169-77.
7. Tripathi Brahma Nanda, Ashtang Hridayam of shrimadvagabhata, Chikitsa sthana, Chaukhamba Sanskrit Pratishtan, Delhi, 2015, Pramehachikitsadhyaya-12/17-18:717
8. Chakapanidatta, Charaka Samhita of Agnivesha, Chikitsasthana. Chaukhamba Subharti Prakashan, Varansi, 2014, Pramehachikitsitam Sashtodhyaya, 6/26-27:447
9. Susruta Samhita (edited with Ayurveda Tattva Sandipika Hindi Commentary, Scientific Analysis, Notes etc. by Kaviraj Ambikadutta Shastri forwarded by Dr. P.M. Mehta), part 1, Chikitsa sthan, 11/8, reprint edition, published by Chaukhambha Sanskrit Sansthan, Varanasi., 2005; 60.
10. DH Colonel, Text book of Ayurvedic Useful plants o India, Second Edition, Asiatic Publication House, Delhi, 2006; 215:169-170.
11. Indian Medicinal Plants, Vol-8, published by Indian Council of Medical Research New Delhi, 2009. 23:312-314
12. Srinivasan K. Plant foods in the management of diabetes mellitus: spices as beneficial antidiabetic food adjuncts. International journal of food sciences and nutrition. 2005 Jan 1;56(6):399-414.
13. Arun N, Nalini N. Efficacy of turmeric on blood sugar and polyol pathway in diabetic albino rats. Plant Foods for Human Nutrition. 2002 Mar 1;57(1):41-52.
14. Babu PS, Srinivasan K. Amelioration of renal lesions associated with diabetes by dietary curcumin in streptozotocin diabetic rats. Molecular and cellular biochemistry. 1998 Apr 1;181(1):87-96.
15. Bhat SD, Ashok BK, Rabinarayan. Exploring the concept of Vacha (*Acorus calamus* Linn.) shodhana in Ayurveda. Int J Res Ayurveda Pharm. 2012;3:341-4.
16. Maji Jayanta Kumar, Sohim, Shukla VJ: Application of multivariate curve resolution alternating least square (MCR-ALS) to the study of Trikarshika formulation. International Journal of Pharmacy and Pharmaceutical Sciences 2016, 8 (3):238-243.
17. Jayanta Kumar Maji. Near-Infrared Spectroscopy: A potential new mean of assessing multicomponent polyherbal formulation on way before and after extraction. International Journal of Pharmacy and Pharmaceutical Sciences 2017; 9(5) :121-129.
18. Eichler HG, Korn A, Gasic S, Pirson W, Businger J. The effect of a new specific α -amylase inhibitor on post-prandial glucose and insulin excursions in normal subjects and Type 2 (non-insulin-dependent) diabetic patients. Diabetologia. 1984 Apr 1;26(4):278-81.
19. Etxeberria U, de la Garza AL, Campión J, Martinez JA, Milagro FI. Antidiabetic effects of natural plant extracts via inhibition of carbohydrate hydrolysis enzymes with emphasis on pancreatic alpha amylase. Expert opinion on therapeutic targets. 2012 Mar 1;16(3):269-97.
20. Lekshmi PC, Arimboor R, Raghu KG, Menon AN. Turmerin, the antioxidant protein from turmeric (*Curcuma longa*) exhibits antihyperglycaemic effects. Natural product research. 2012 Sep 1;26(17):1654-8.
21. Kumar S, Narwal S, Kumar V, Prakash O. α -glucosidase inhibitors from plants: A natural approach to treat diabetes. Pharmacognosy reviews. 2011 Jan 1;5(9):19.
22. Siddhinanadana Mishra, "Harmekhala", Chaukhambha Publication, Varansi, Edition-1st 2013. 5/161, pp-166
23. Ramtejpandeyana, Bhavaprakash nighantoo, 3rd edition, Varansi: Chaukhamba Sanskrita Pratishtana, 2003. Takra varga 15/1-2, pp-304
24. Thomas Lijio and P. Rajeev, Turmeric - Indian Institute of Spices Research; Turmeric - Extension Pamphlet, ICAR, Kozhikode publisher; November 2015, pp-9-10; Access by <http://www.spices.res.in/pdf/package/turmeric.pdf> (12-01-2017)
25. Govind Das. Bhaisajyaratnavali, Ambikadatta sastry editor. 15th edition; Varansi: Chaukhamba Sanskrita samsthana; 2001. Vatavyadhichikitsa-26/447,pp.-404
26. Tripathi Jagdishprasad, Chakradatta of Chakarpanidatta. 5th edition Varansi: Chaukhamba Sanskrit series office, 1983; Vatavyadhichikitsa 22/287:211.
27. Thalapaneni NR, Chidambaram KA, Ellappan T, Sabapati ML, Mandal SC. Journal of complementary and Integrative Medicine 2008; 5(1): 1-10.
28. Heidari R, Zareae S, Heidarizadeh M. Extraction, purification, and inhibitory effect of alpha-amylase inhibitor from wheat (*Triticum aestivum* Var. Zarrin). Pakistan Journal of Nutrition. 2005;4(2):101-5.
29. Andrade-Cetto A, Becerra-Jiménez J, Cárdenas-Vázquez R. Alfa-glucosidase-inhibiting activity of some Mexican plants used in the treatment of type 2 diabetes. Journal of ethnopharmacology. 2008 Feb 28;116(1):27-32.
30. Matsuura H, Asakawa C, Kurimoto M, Mizutani J. α -Glucosidase inhibitor from the seeds of balsam pear (*Momordica charantia*) and the fruit bodies of *Grifola frondosa*. Bioscience, biotechnology, and biochemistry. 2002 Jan 1;66(7):1576-8.
31. Tietz, N.W. In: Burtis, C.A., Ashwood, E.R. (Eds.), Tietz Textbook of Clinical Chemistry, third ed. Saunders W.B., 1999, 1;48(1):750-778.
32. Tieting CA, Woods K, Zhang R, Brastianos HC, Brayer GD, Andersen RJ, Withers SG. The Search for Novel Human Pancreatic α -Amylase Inhibitors: High-Throughput Screening of Terrestrial and Marine Natural Product Extracts. ChemBioChem. 2008 Feb 15;9(3):433-8.
33. Sudha P, Zinjarde SS, Bhargava SY, Kumar AR. Potent α -amylase inhibitory activity of Indian Ayurvedic medicinal plants. BMC complementary and alternative medicine. 2011 Jan 20;11(1):5.
34. Paul T, Banerjee S. Invitro evaluation of α -amylase inhibitory activity & antioxidant potential of *Pteris vittata* L. with special reference to its HPTLC profile. International Journal of Pharma and Bio Sciences. 2013:494-503.

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